

Distribution of phytoliths in plants: a review

Rajat Sharma, Vinod Kumar & Rakesh Kumar

To cite this article: Rajat Sharma, Vinod Kumar & Rakesh Kumar (2019) Distribution of phytoliths in plants: a review, *Geology, Ecology, and Landscapes*, 3:2, 123-148, DOI: [10.1080/24749508.2018.1522838](https://doi.org/10.1080/24749508.2018.1522838)

To link to this article: <https://doi.org/10.1080/24749508.2018.1522838>



© 2018 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group on behalf of the International Water, Air & Soil Conservation Society (INWASCON).



Published online: 25 Sep 2018.



[Submit your article to this journal](#)



Article views: 1273



[View related articles](#)



[View Crossmark data](#)



Citing articles: 8 [View citing articles](#)

Distribution of phytoliths in plants: a review

Rajat Sharma, Vinod Kumar and Rakesh Kumar

Department of Botany, DAV University, Jalandhar, India

ABSTRACT

Phytoliths are ergastic siliceous substances present abundantly within intercellular spaces as well as inside the cells of numerous plants. Being made up of silica, they are nondegradable and hence found preserved as microfossils in various substrata. This property of phytoliths extends its significance in the field of paleobotany, geology, and archaeology. Soil analysis for eking out phytoliths has been often exercised for reconstructing paleophytogeography of extinct grasslands. Plants accumulate silica and convert them into crystals when their concentration reaches certain maxima. Deciphering the types of phytoliths at historically rich sites, we can predict the culture of ancient farming. Finding such peerless opulence of phytoliths in paleobotany, the present review was designed to compile the data of phytoliths in various plants. The data compiled in this review will help researchers tracing the link between phytoliths and its source plant at the target sites.

ARTICLE HISTORY

Received 18 April 2018

Accepted 10 September 2018

KEYWORDS

Phytoliths; Pteridaceae; dry ashing method

Introduction

Silica is important for the growth and development of the plants (Agarie, Agata, Uchida, Kubota, & Kaufman, 1996). It is absorbed through roots in the form of monosilicic acid through the water conducting tissue, i.e., xylem (Blackman and Parry, 1968). Many plant species deposit silica in their intercellular or extracellular location in the cells. These silica bodies are called phytoliths. They are found deposited in both over and between leaf veins, culms, and in epidermis of inflorescence and rarely in seeds (Zsuzsa et al., 2014). Phytolith has been used for various types of mineral deposition that may be siliceous and calcareous particles that show a great difference in their structure and taxonomic attributes (Shakoor & Bhat, 2014). These have been extracted from many different plant organs such as leaves, stems, inflorescence, seeds, and roots. Current knowledge suggests that phytoliths are restricted to the vascular plants, with high production levels and common family-specific forms occurring in the pteridophytes (tree ferns and horsetails), basal angiosperms (magnolias), and monocotyledons (particularly the grasses and sedges), eudicots. These are inorganic and thus resistant to the forces of decay that cause the destruction of other types of plant materials, and they survive in a well-preserved state over long periods of time. These have been found to be advantageous for the growth and development of many plants (Matoh et al., 1986). It also helps the plants to overcome various biotic and abiotic stresses

(Epstein, 1999; Ma and Yamaji, 2006). The existence of phytoliths in the plant tissues has structural and protective role against fungi, insects, and herbivores (Shakoor et al., 2014). Silica bodies are abundant and morphologically distinct as well as more durable in soil for 1000 years even after the death and decay of the plants (Piperno & Pearsall, 1998). The structural hierarchy and durability of this element in the form of phytoliths in different parts of the plant (Chauhan, Tripathi, Kumar, & Kumar, 2011) as well as in the soil has proven the possibilities to solve the archaeological problems and to identify the crop plants which were grown in our past (Twiss 1987). The archeological and paleoecological significance of phytoliths has researched under the various names that are opal phytoliths, silica phytoliths, silica cells, plant opal, biogenic opal, or simply phytoliths (Albert et al., 2001). Phytoliths are present in various shape and sizes depending upon the location of its deposition and the age of the plant. Plant does not use the silica for any of its metabolic processes and deposit them as silica gel. As the concentration of silica gel increases, it gradually crystallizes into a solid silt-sized particle. When plant dies, the phytoliths are released into the soil (Shakoor & Bhat, 2014). Numerous angiosperms, gymnosperms, and pteridophytes produce large quantity of phytoliths. Among the pteridophytes species of Equisetaceae, there are heavy silicon accumulators which use them as nutrient (Chen et al., 1969). Silica may get deposited in any of the cells of dermal,

ground, and the vascular tissues. Hyper-accumulation of silicon in plants helps them for growing under biotic and abiotic stresses like heavy metal, drought, salinity, and pathogens (Ma and Yamaji, 2006). Among recent advances in plant identification, phytoliths have got substantive importance in the identification of different taxa at various levels of hierarchy. Many taxa in family Poaceae are characterized by the phytoliths with specific morphological characteristics; hence, they extend their significance in taxonomic as well. When the plants die, phytoliths are released from the plant tissues, and released phytoliths from the plant tissues become the microfossil of that plant. These microfossils are formed in different plant parts, depending on the taxa and most commonly within epidermal cells of aerial structures but also sometimes within wood, mesophyll, and roots. Silica deposition can be actively controlled by plants through physiological mechanisms and in relation to specific cells and passive potentially leading to silicification of a more variable range of cells and areas of plant tissue. Areas of epidermal tissue can become silicified, and these conjoined phytoliths are commonly referred to as silica skeletons (Ryan et al., 2014). Production of silica in the bark is greater than the wood; phytoliths in wood have long since been recorded in the frame of wood anatomical research (Collura & Neumann, 2017). However, many plant species do not produce phytoliths but an important factor for understanding phytolith production is that patterns of solid silicon accumulation, together with the placement of the hardened deposits in specific tissues and cells of plants, are quite similar in plant species and their most closely related taxa regardless of the environmental conditions of growth (Shakoor & Bhat, 2014). The initial interest in phytolith type was focused on their diagnostic potential in plant taxonomy and phytoliths were observed *in situ*. Study of phytoliths morphology in grasses and modern plants can be used for the identification of botanical remains in fossil records (Esteban et al., 2016). However, the physiological role of silicon in vascular plants is still an open subject; silicon may act as an essential or a beneficial element depending on the species. So, the phytolith analysis can be a valuable tool in the study of the vegetation history (Carnelli, Theurillat, & Madella, 2004). The present review was designed to show the distribution of different phytolith types present in various plant species.

Collection of data

The data of phytoliths were collected from 21 research articles from 1978 to 2017 and presented in Table 1. Figure 1 showed the number of families in which phytolith analysis has been reported and from the analyzed data it was found that Pteridaceae (156 plant species) followed by Poaceae (140 plant species) and Fabaceae (29 plant species) showed maximum number of plant species in which phytolith analysis has been done. The various methodological approaches adapted by researchers for the phytolith analysis were shown in Figure 2. From the data, it was found that dry ashing method was followed by wet oxidation method, and wet ashing methods are mainly applied for the digestion of samples to carry out the phytolith analysis.

Future directions

It has been more than 150 years since the phytolith research began in Germany which interpreted the shape of silica bodies formed in plants (Piperno, 1988, 2006; Struve, 1835). Since then substantive phytolith research has been progressed in various field of science. The archaeologists used phytolith to predict early human agritarian contributions to understand human past. The extensive research is required to understand the isotopic depositions of phytoliths (Hart, 2016). Phytolith analysis of top soil helps in reconstructing paleological environments of various proxies (Wu et al., 2018). However, lack of standard procedures is the major stumbling block to exploit full potential of phytolith research in wider areas pertaining to development of reliable models in archeological studies (Zurro, García-Granero, Lancelotti, & Madella, 2016). There is compelling necessity of describing additional geographical areas for available phytolith diversity (Zurro et al., 2016). Phytolith carbon sequestration presents mainly to the carbon cycle, and hence it is important to integrate plant-soil silica cycling in biogeochemical cycling of carbon. Studies with terrestrial biogeochemical cycles should pursue more clearly about the plant-soil silica-carbon modeling to drive the biogeochemical carbon sequestration (Song, Liu, Strömberg, Yang, & Zhang, 2017). Sequestration of carbon in phytoliths is considered as stable carbon sink process in terrestrial ecosystems to reduce the impact of



Table 1. Distribution of phytoliths in different plant species.

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Angiosperms						
Areaceae	Areaceae	Wet oxidation technique	<i>Sabal minor</i>	Leaf stem fruit	Spherical, elliptical, hat, conical shaped	Piperno(2006)
Areaceae	Areaceae	Wet oxidation technique	<i>Sabal etonia</i>	Leaf stem fruit	Spherical, elliptical, hat, conical shaped	Piperno (2006)
Areaceae	Areaceae	Wet oxidation technique	<i>Serenoa repens</i>	Leaf stem fruit	Spherical, elliptical, hat, conical shaped	Piperno (2006)
Cyperaceae	Cyperaceae	Wet oxidation method	<i>Carex curvula</i>	Leaf fruit seed	Hat shaped, mesophyll	Piperno (2006)
Cyperaceae	Cyperaceae	Wet oxidation method	<i>Cyperus polystachyos</i>	Leaf fruit seed	Hat shaped, mesophyll	Piperno (2006)
Cyperaceae	Cyperaceae	Wet oxidation method	<i>Schoenus nigricans</i>	Leaf fruit seed	Hat shaped, mesophyll	Piperno (2006)
Liliaceae	Liliaceae	Wet oxidation technique	<i>Yucca aloifolia</i>	Leaf root stem	Rondel saddle	Thorne & Kishino (2002)
Miarantaceae	Miarantaceae	Wet oxidation technique	<i>Maranta arundinacea</i>	Seed fruit	Conical, spherical, nodulose	Piperno (2006)
Poaceae	Poaceae	Wet oxidation technique	<i>Andropogon scoparius</i>	Leaf	Polylobate, rondel, elongate	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Cenchrus longispinus</i>	Leaf	Polylobate, rondel, elongate	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Festuca scarbriculims</i>	Leaf	Polylobate, rondel, elongate	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Nardus stricta</i>	Leaf	Polylobate, rondel, elongate	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Andropogon glomeratus</i>	Leaf	Dumbell	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Andropogon ternarius</i>	Leaf	Dumbell two-horned tower	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Anthaenatia rufa</i>	Leaf	Dumbell cross	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Aristida desmantha</i>	Leaf	Dumbell rondel two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Arundinaria longifolia</i>	Leaf	Long saddle short saddle two-horned spool	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Arundinaria gigantea</i>	Leaf	Long saddle short saddle two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Avena sativa</i> L.	Leaf	Rondel	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Cenchrus incertus</i>	Leaf	Dumbell cross two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Chasmanthium laxum</i>	Leaf	Dumbell cross two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Chasmanthium ornithorhynchum</i>	Leaf	Dumbell rondel cross	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Ctenium aromaticum</i>	Leaf	Dumbell flat tower two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Dactyloctenium aegyptium</i>	Leaf	Short saddle	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Distichlis spicata</i>	Leaf	Short saddle two-horned tower spool	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Eragrostis oxylepis</i>	Leaf	Short saddle two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Eragrostis cilianensis</i>	Leaf	Short saddle two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Erianthus strictus</i>	Leaf	Short saddle two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Eustachys petraea</i>	Leaf	Shorty saddle two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Leersia oryzoides</i>	Leaf	Dumbell two-horned tower	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Panicum amarum</i>	Leaf	Dumbell spool	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Panicum dichotomiflorum</i>	Leaf	Dumbell cross two-horned tower	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Panicum hemitomom</i>	Leaf	Dumbell cross	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Panicum verrucosum</i>	Leaf	Dumbell	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Panicum virgatum</i>	Leaf	Dumbell two-horned	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Phragmites australis</i>	Leaf	Short saddle flat tower	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Poa annua</i>	Leaf	Rondel flat tower	Korolük (2010)
Poaceae	Poaceae	Wet oxidation technique	<i>Saccharum officinarum</i>	Leaf	Dumbell	Korolük (2010)

(Continued)



Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Poaceae		Wet oxidation technique	<i>Setaria</i> sp.	Leaf	Dumbbell	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Sorghastrum nutans</i>	Leaf	Dumbbell cross two-horned	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Sorghum halepense</i>	Leaf	Dumbbell cross	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Sorghum halepense</i>	Leaf	Dumbbell cross	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Spartina alterniflora</i>	Leaf	Short saddle flat tower	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Spartina patens</i>	Leaf	Short saddle dumbbell	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Sporobolus virginicus</i>	Leaf	Flat tower two-horned tower spool	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Uniola paniculata</i>	Leaf	Flat two-horned tower spool	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Zizaniopsis miliacea</i>	Leaf	Dumbbell two-horned tower	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Oryza sativa</i>	Inflorescence	Polylobate, rondel, elongate	Korolük (2010)
Poaceae		Wet oxidation technique	<i>Zea mays</i>	Corn leaf husk tassel	Bilobate, rondel, saddle, irregular, elongate	Korolük (2010)
Poaceae		Ultrasound water bath & wet oxidation technique	<i>Calamagrostis villosa</i>	Leaf culm	Bilobate, rondel, saddle, irregular, elongate	Cameli et al. (2001)
Poaceae		Ultrasound water bath & wet oxidation technique	<i>Festuca halleri</i>	Leaf culm	Bilobate, rondel, saddle, irregular, elongate	Cameli et al. (2001)
Poaceae		Ultrasound water bath & wet oxidation technique	<i>Festuca puccinelli</i>	Leaf culm	Bilobate, rondel, saddle, irregular, elongate	Cameli et al. (2001)
Poaceae		Ultrasound water bath & wet oxidation technique	<i>Festuca scabriculumis</i>	Leaf culm	Bilobate, rondel, saddle, irregular, elongate	Cameli et al. (2001)
Poaceae		Ultrasound water bath & wet oxidation technique	<i>Poa alpina</i>	Leaf	Bilobate, rondel, saddle, irregular	Cameli et al. (2001)
Poaceae		Ultrasound water bath & wet oxidation technique	<i>Nardus stricta</i>	Leaf	Bilobate, rondel, saddle, irregular, elongate	Cameli et al. (2001)
Poaceae		Wet oxidation technique	<i>Sorghum vulgare</i>	Leaf	Rondel, saddle, irregular, elongate	Korolük (2010)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Bambusa</i> sp.	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Li et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>B. rutila</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>B. multiplex</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>B. multiplex</i> cv. <i>Changye</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>B. multiplex raeuschel</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>B. alphonsekarri</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>B. glaucescens</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>H. tranguillans.shiroshima</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sh. kumasasa</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sh. chinensis nakai</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sh. chinensis nakai cv. Jimao</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>S. yashadake f. kimmei</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>S. yashadake makino</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>S. yashadake f. ogon</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys prominens</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys vivax</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys heterocycla taokiang</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys heterocycla</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys incarnata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys bambusoides</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys bambusoides cv. huayehuagan</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys bambusoides.castillonis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys Nigra</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys aureosulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys aureosulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys.aureosulcata.spectabilis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys.sulphurea.viridis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys sulphurea viridisulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys sulphurea</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys houzeauana</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys ventricosa</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys ventricosa cv huangganlucao</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys ventricosa cv. Luanhuangcao</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys ventricosa cv. huangjin</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys arcana.luteosulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys propinqua</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys vivax.aureocaulis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys heterocycla.gracilis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys nigra.henonis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys dulcis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys panvifolia</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys violascens cv. Xiye</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys nviolascens cv. Jianye</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys violascens cv. viridisulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys sulphurea</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys houzeauana</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys Ventricosa</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys ventricosa cv huangganlucao</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys ventricosa</i> cv. <i>Luganhuangcao</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys ventricosa</i> cv. <i>huangjin</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys arcana.luteosulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys propinqua</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys vivax.aureocaulis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys heterocycla.gracilis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys nigra.henonis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys edulis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys parvifolia</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys violascens</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys violascens</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys viridisulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys flavistriatus</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys panggan</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys anhuiensis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys lavivaginis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys violascens</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys bambusoides</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys aureosulcata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Phyllostachys</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>I. acutiligulata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Indosasasinica</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sasaellarootsisk</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sasaellaauricoma</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Chimonobambusaquadrangularis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pleioblastuskongosamensis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pleioblastus hisauchii</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pleioblastus simony</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pleioblastus nearis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pleioblastus angustifolius</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pseudosasa amabilis</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pseudosasa japonica</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Pseudosasa tsutsumiana</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Acidosasagigantean</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Oligostachyum</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Oligostachyum rubricum</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sasa argenteostriata</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sasapygmaea</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Sasa auricoma</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Indosasadecoratus</i>	Leaf	Rondel, saddle, irregular, elongate, bilobate, flat	Beilei et al. (2014)
Poaceae		Oven-dried & Walkley-Black type digest technique	<i>Bromus inermis</i>	Leaf	Polylobate, elongate, plate, rondel, bilobate	Beilei et al. (2014)
Musaceae		Rapid microwave digestion technique	<i>Musa paradisiaca</i>	Leaf	Irregular shape	Lentfer et al. (2002)
Heliconiaceae		Rapid microwave digestion technique	<i>Heliconia</i> spp.	Seed leaf	Irregular shape	Lentfer et al. (2002)

(Continued)



Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Restionaceae		Dry ash technique	<i>Restio triticeus</i>	Stem	Parallelepipedal elongate, angulate, scrobiculate	Novello et al. (2017)
Orchidaceae		Rapid microwave digestion technique	<i>Orchid</i> spp.	Leaf	Conical	Lentfer et al. (2002)
Aceraceae		Wet oxidation technique	<i>Acer saccharum</i> , <i>Acer negundo</i>	Leaf	Lanceolate, spherical, elliptical	Piperno (2006)
Asteraceae		Wet oxidation technique	<i>Helianthus annuus</i>	Leaf	Tracheid, mesophyll	Piperno (2006)
Asteraceae		Sonication & dried technique	<i>Aspilia mosambicensis</i>	Leaf	Cylindrical Scrobiculate	Mercader, Bennett, Esselmont, Simpson, & Walde (2009)
Asteraceae		Sonication & dried technique	<i>Brachylaena</i> spp.	Stem leaf	Tracheid, mesophyll cylindrical scrobiculate	Mercader et al. (2009)
Asteraceae		Sonication & dried technique	<i>Pleiotaxis</i> spp.	Leaf	Cylindrical scrobiculate	Mercader et al. (2009)
Asteraceae		Sonication & dried technique	<i>Aspilia</i> spp.	Leaf	Tracheid, mesophyll cylindrical scrobiculate	Mercader et al. (2009)
Asteraceae		Sonication & dried technique	<i>Vernonia amygdalina</i>	Leaf stem	Tracheid, mesophyll, cylindrical scrobiculate	Mercader et al. (2009)
Anacardiaceae		Dry and wet ashing technique	<i>Lannea fruticosa</i>	Wood	Elongate, scalariform, elongate, vesicular, blocky, irregular tabular, irregular	Collura & Neumann (2017)
Anacardiaceae		Dry and wet ashing technique	<i>Lannea acida</i>	Wood	Elongate, scalariform, elongate, vesicular, blocky, irregular tabular, irregular	Collura & Neumann (2017)
Sapotaceae		Dry and wet ashing technique	<i>Vitellaria paradoxa</i>	Wood	Elongate, scalariform, elongate, vesicular, blocky, irregular tabular, irregular	Collura & Neumann (2017)
Betulaceae		Wet oxidation technique	<i>Carpinus Carolina</i> , <i>Ostrya virginiana</i>	Leaf	Lanceolate spherical elliptical	Piperno (2006)
Cucurbitaceae		Wet oxidation technique	<i>Cucurbita pepo</i>	Fruit seed	Lanceolate, spherical elliptical	Piperno (2006)
Dilleniaceae		Wet oxidation technique	<i>Curatella americana</i> , <i>Tetracera volubilis</i>	Leaf wood	Elliptical lanceolate, spherical	Piperno (2006)
Ericaceae		Wet oxidation technique	<i>Rhododendrum ferrugineum</i> , <i>Vaccinium myrtillus</i>	Leaf	Lanceolate, spherical elliptical	Piperno (2006)
Ericaceae		Wet oxidation technique	<i>Euclaea crispa</i>	Leaf	Lanceolate, spherical elliptical	Mercader et al. (2009)
Proteaceae		Dry ash technique	<i>Diospyros</i> spp.	Leaf	Lanceolate, spherical elliptical	Mercader et al. (2009)
Proteaceae		Dry ash technique	<i>Leucadendron salignum</i>	Leaf	Rondelet, trapeziform, cubic/rectangular	Novello et al. (2017)
Proteaceae		Dry ash technique	<i>Leucadendron spissifolium</i>	Leaf	Rondelet, trapeziform, cubic/rectangular	Novello et al. (2017)
Proteaceae		Dry ash technique	<i>Leucospermum cuneifolia</i>	Leaf	Rondelet, trapeziform, cubic/rectangular	Novello et al. (2017)
Proteaceae		Dry ash technique	<i>Protea nerifolia</i>	Leaf	Rondelet, trapeziform, cubic/rectangular	Novello et al. (2017)
Proteaceae		Dry ash technique	<i>Protea tena</i>	Leaf	Rondelet, trapeziform, cubic/rectangular	Novello et al. (2017)
Vitaceae		Dry ash technique	<i>Rhoicissus tomentosa</i>	Leaf	Rondelet, trapeziform, cubic/rectangular	Novello et al. (2017)
Euphorbiaceae		Wet oxidation technique	<i>Manihot esculenta</i>	Leaf fruit seed	Lanceolate, spherical elliptical	Piperno (2006)
Fagaceae		Wet oxidation technique	<i>Quercus macrocarpa</i> , <i>Quercus faevis</i>	Leaf	Lanceolate, spherical elliptical	Piperno (2006)
Juglandaceae		Wet oxidation technique	<i>Carya ovata</i> , <i>Juglans nigra</i>	Leaf	Lanceolate, spherical elliptical	Piperno (2006)
Moraceae			<i>Morus rubra</i>	Leaf bark	Hook shaped	Geiset et al. (1973)
Rubiaceae			<i>Coffea arbica</i>	Leaf wood	Rondelet	Thorn (2008)
Fabaceae		Sonication & dried technique	<i>Albizia anthelmintica</i>	Leaf stem	Block with lacunose/slightly scrobiculate texture	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Lonchocarpus capassa</i>	Leaf	Blocky cavate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Dolichos kilimandscharicus</i>	Leaf	Blocky comiculate	Mercader et al. (2009)

(Continued)



Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Fabaceae		Sonication & dried technique	<i>Lonchocarpus capassa</i>	Stem	Blocky hairy	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Sphenostylis</i> spp.	Leaf	Cylindrical	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Ptilostigma thomningii</i>	Leaf	Cylindrical bulbous	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Afzelia quanzensis</i>	Leaf	Epidermal laminate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Pterocarpus angolensis</i>	Stem	Globular echinate large	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Afzelia quanzensis</i>	Leaf	Globular psilate large	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Pterocarpus angolensis</i>	Stem	Globulose bisected	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Sphenostylis</i> spp.	Stem	Lenticular concave/ convex	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Cajanus cajan</i>	Leaf	Tabular crenate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Dolichos kilimandscharicus</i>	Stem	Tabular scrobiculate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Elephantorrhiza goetzei</i>	Leaf	Tabular thick lacunate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Pterocarpus angolensis</i>	Leaf	Tabular thick lacunate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Pterocarpus tinctorius</i>	Leaf	Tabular thick sinuate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Albizia gummifera</i>	Leaf	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Brachystegia</i> spp	Stem	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Dalbergiella nyasae</i>	Stem	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Acacia tortilis</i>	Leaf	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Acacia schweinfurthii</i>	Leaf	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Pterocarpus tinctorius</i>	Stem	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Acacia</i> spp	Leaf	Tabular thick lacunate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Brachystegia</i> spp	Leaf	Tabular thick lacunate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Aeschynomene</i> spp	Leaf	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Securidaca Longipedunculata</i>	Stem	Tabular trapezoid	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Erythrophileum suaveolens</i>	Leaf	Tabular thick lacunate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Brachystegia</i> spp	Leaf	Tabular thick lacunate	Mercader et al. (2009)
Fabaceae		Sonication & dried technique	<i>Brachystegia boehmii</i>	Stem	Tabular thick lacunate	Mercader et al. (2009)
Rosaceae		Sonication & dried technique	<i>Ficus</i> spp.	Leaf	Cylindrical scrobiculate	Mercader et al. (2009)
Rosaceae		Sonication & dried technique	<i>Parinari</i> spp	Leaf stem	Globular facetate	Mercader et al. (2009)
Rosaceae		Sonication & dried technique	<i>Pouzolzia mixta</i>	LeafStem	Cylindrical scrobiculate	Mercader et al. (2009)
Rosaceae		Sonication & dried technique	<i>Ensete ventricosum</i>	Stem	Globular facetate	Mercader et al. (2009)
Rosaceae		Sonication & dried technique	<i>Ziziphus mucronata</i>	Stem	Cylindrical scrobiculate	Mercader et al. (2009)
Celastraceae		Dry ash technique	<i>Pterocelastrus tricuspidatus</i>	Leaf	Parallelepipedal elongate, angulate, scorbiculate	Novello et al. (2017)
Ulmaceae		Wet oxidation technique	<i>Celtis occidentalis, Ulmus americana</i>	Fruit seed bark	Sinuate spherical	Thorn (2008)
Gymnosperms		Wet oxidation technique	<i>Abies balsamea</i>	Needle foliage, wood	Flat spiny granulated	Piperno (2006)
Pinaceae		Wet oxidation technique	<i>Abies fraseri</i>	Needle foliage, wood	Flat spiny granulated	Klein and Geis (1978)
Pinaceae		Wet oxidation technique	<i>Abies grandis</i>	Needle foliage, wood	Flat spiny granulated	Piperno (2006)
Pinaceae		Wet oxidation technique	<i>Pinus strobus</i>	Needle foliage, wood	Flat spiny granulated	Piperno(2006)
Pinaceae		Wet oxidation technique	<i>Podocarpus neriifolia</i>	Needle foliage, wood	Flat spiny granulated	Piperno(2006)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pinaceae		Ultrasonic bath technique	<i>Carex lanceolata</i>	Needle foliage, wood	Flat spiny granulated	Yang, Wu, Liu, Chen, and Zhou (2018)
Pinaceae		Ultrasonic bath technique	<i>Artemisia eriopoda</i>	Needle foliage, wood	Flat spiny granulated	Yang et al. (2018)
Pinaceae		Ultrasonic bath technique	<i>Lespedeza bicolor</i>	Needle foliage, wood	Flat spiny granulated	Yang et al. (2018)
Pinaceae		Ultrasonic bath technique	<i>Pinus tabulaeformis</i>	Needle foliage, wood	Flat spiny granulated	Yang et al. (2018)
Taxaceae		Dry ashing technique	<i>Taxus baccata</i> , <i>Taxus cuspidate</i>	Needle, wood	Flat spiny granulated	Camelli et al. (2004)
Taxodiaceae		Wet oxidation technique	<i>Taxodium distichum</i>	Needle wood	Flat spiny granulated	Piperno (2006)
Araucariaceae		Dry ashing technique	<i>Araucaria araucana</i>	Needle, foliage	Flat spiny granulated	Blinnikov (2005)
Cupressaceae		Dry ashing technique	<i>Juniperus communis</i> , <i>Juniperus nana</i>	Needle wood	Flat spiny granulated	Klein and Geis (1978)
Pinaceae		Dry ashing technique	<i>Picea jezoensis</i>	Wood needle	Bilobate short cell	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Abies nephrolepis</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Pinus koraiensi</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Larix olgensis</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Rosa marretii</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Solidago virgaurea</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Calamagrostis angustifolia</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Beckmannia syzigachne</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Pinaceae		Dry ashing technique	<i>Arthraxon hispidus</i>	Wood needle	Saddle rondel rectangle dentate	Yang et al. (2018)
Epacridaceae		Low-temperature acid extraction technique	<i>Dracophyllum scoparium</i>	Leaf wood	Rondel	Thorn (2008)
Scrophulariaceae		Dry ashing technique	<i>Hebe elliptica</i>	Leaf	Saddle rondel rectangle dentate	Yang et al. (2018)
Rosaceae		Low-temperature acid extraction (Jones & Milne 1966)	<i>Acaena anserinifolia</i>	Leaf	Conical elongated trapezoid globular	Thorn (2008)
Boraginaceae		Low-temperature acid extraction (Jones & Milne 1966)	<i>Myosotis capitata</i>	Leaf	Conical elongated trapezoid globular	Thorn (2008)
Pteridophytes	Equisetaceae	Hot bath & acid treatment technique	<i>Equisetum</i>	Stem leaf	Elongate echinate, cylindrical polylobate, acicular, papillate, stomata, stomata	Stromberg et al. (2002)
Lycopodiaceae		Dry ashing technique	<i>Lycopodium clavatum</i>	Whole plant	Smooth or granular plates with projecting margins	Chauhan et al. (2011)
Lycopodiaceae		Wet oxidation technique	<i>L. japonicum</i> Thunb	Aerial branch	Smooth or granular plates with projecting margins	Mazumdar (2011)
Lycopodiaceae		Wet oxidation technique	<i>Palhinhaea cernua</i>	Aerial	Smooth or granular plates with projecting margins	Mazumdar (2011)
Lycopodiaceae		Wet oxidation technique	<i>Huperzia selago</i>	Aerial	Smooth or granular plates with projecting margins	Mazumdar (2011)
Selaginellaceae		Wet oxidation technique	<i>Selaginella bryopteris</i>	Whole plant	Elongated epidermal bodies with small cone like projections	Mazumdar (2011)
Selaginellaceae		Wet oxidation technique	<i>Selaginella inaequalifolia</i>	Aerial shoot	Elongated epidermal bodies with small cone like projections	Mazumdar (2011)
Selaginellaceae		Wet oxidation technique	<i>Selaginella involvens</i>	Leaf	Elongated epidermal bodies with small cone like projections	Mazumdar (2011)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
	Selaginellaceae	Wet oxidation technique	<i>Selaginella monospora</i>	Aerial shoot	Elongated epidermal bodies with small cone like pro-Elongated epidermal bodies with small cone like projections	Mazumdar (2011)
	Selaginellaceae	Wet oxidation technique	<i>Selaginella pentagona</i>	Aerial shoot	Elongated epidermal bodies with small cone like projections	Mazumdar (2011)
	Selaginellaceae	Wet oxidation technique	<i>Selaginella plana</i>	Aerial shoot	Elongated epidermal bodies with small cone like projections	Mazumdar (2011)
	Selaginellaceae	Wet oxidation technique	<i>Selaginella tenera</i>	Aerial shoot	Elongated epidermal bodies with small cone like projections	Mazumdar (2011)
	Selaginellaceae	Wet oxidation technique	<i>Selaginella chrysocaulos</i>	Aerial shoot	Elongated epidermal bodies with small cone like projections	Mazumdar (2011)
	Selaginellaceae	Wet oxidation technique	<i>Selaginella velutina</i>	Leaf	Elongated epidermal bodies with small cone like projections	Piperno (2006)
	Isotaceae	Wet oxidation technique	<i>Isôetes coromandelina</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
			<i>Isôetes weberi</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Iriarte & Paz (2009)
	Ophioglossaceae	Wet oxidation technique	<i>Botrychium virginianum</i>	Leaf, spike	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
			<i>Helminthostachys zeylanica</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
			<i>Ophioglossum reticulatum</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
	Marattiaceae	Wet oxidation technique	<i>Angiopteris evecta</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
	Hymenophyllaceae	Wet oxidation technique	<i>Hymenophyllum</i> sp.	Leaf	Epidermal cells like angular, polyhedral bodies	Piperno (2006)
	Hymenophyllaceae	Wet oxidation technique	<i>Trichomanes</i> sp.	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
	Gleicheniaceae	Wet oxidation technique	<i>Dicranopteris linearis</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
			<i>Gleichenia gigantean</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
			<i>Gleichenia microphylla</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
			<i>Lygodium flexuosum</i>	Whole plant	Smooth or granular plates with projecting margins	Chauhan et al. (2011)
	Marsileaceae		<i>Marsilea ancylopoda</i>	Leaf		Iriarte and Paz (2009)
	Cyatheaceae	Dry ashing technique	<i>Cyathea gigantea</i>	Whole plant	Smooth or granular plates with projecting margins	Chauhan et al. (2011)
	Cyatheaceae	Wet oxidation technique	<i>Cyathea spinulosa</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
	Lindsaeaceae	Wet oxidation technique	<i>Lindsaea odorata</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
	Lindsaeaceae	Wet oxidation technique	<i>Sphenomeris chinensis</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
	Demstaedtiaceae	Wet oxidation technique	<i>Hypolepis punctata</i>	Leaf	Epidermal cells like angular, polyhedral bodies	Mazumdar (2011)
	Pteridaceae	Wet ashing technique	<i>Acrostichum danaeifolium</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
	Pteridaceae	Wet ashing technique	<i>Actiniopteris australis</i>	Whole plant	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Chauhan et al. (2011)
	Pteridaceae	Wet ashing technique	<i>Adiantopsis radiata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae		Wet ashing technique	<i>Adiantum abscissum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum adiantoides</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum andicola</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum argutum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum cajennense</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum capillus-junonis</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum capillus-veneris</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum caryotideum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum caudatum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum concinnum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum cordatum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum delicatulum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum dolosum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum formosum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum fruticosum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum glaucescens</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum hispidulum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum humile</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum incertum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum intermedium</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum jordanii</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)

(Continued)



Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum kaulfusii</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum kendalii</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue(2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum krameri</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum latifolium</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum leprieurii</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum lucidum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum lunulatum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum macrophyllum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum malesianum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum mcvaughii</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum multisorum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum oaxacanum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum obliquum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum paraense</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum patens</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum peruvianum</i>	Whole plant	Elliptical shape, tapered ends and undulate sides	Chauhan et al. (2011),
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum petiolatum</i>	Leaf	Elliptical shape, tapered ends and undulate sides	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum philippense</i>	Whole plant	Elliptical shape, tapered ends and undulate sides	Chauhan et al. (2011),
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum phyllitidis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum platyphyllum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum poeppigianum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum poiretii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum polyphyllum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum pulverulentum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum pyramidale</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum raddianum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum reniforme</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum scalare</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum seemanii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum serratodentatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum tenerum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum terminatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum tetraphyllum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Adiantum tomentosum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae		Wet ashing technique	<i>Adiantum trapeziforme</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum trichochlaenum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum tricholepis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum venustum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum villosissimum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum villosum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum vogellii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Adiantum wilsonii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Afropteris repens</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Aleutopteris argentea</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Aleutopteris aurantiacea</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Aleutopteris farinosa</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Ananthacorus angustifolius</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Anetium citrifolium</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae		Wet ashing technique	<i>Anogramma leptophylla</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Anopteris hexagona</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum boyanum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum callifolium</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum latifolium</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum lineatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum mannianum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum minersum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum plantagineum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum reticulatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum brasilianum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Antrophyum obovatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Argyroschosma limitanea</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Aspidotis californica</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)



Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae		Wet ashing technique	<i>Aspidotis densa</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Astrolepis sinuate</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Austrogramme decipiens</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Bommeria hispida</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Ceratopteris pteridoides</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Ceratopteris richardii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Cheilanthes alabamensis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Cheilanthes albomarginata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Cheilanthes angustifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Cheilanthes angustifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Cheilanthes decomposita</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Cheilanthes eatonii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Cryptogramma crispata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Doryopteris ludens</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)



Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae		Wet ashing technique	<i>Doryopteris sagittifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Eriosorus hispidulus</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Haplopteris anguste-elongata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Haplopteris elongata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Haplopteris ensiformis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Haplopteris ensiformis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Haplopteris forestiana</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Haplopteris forestiana</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Hecistopteris pumila</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Hemionitis arifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Hemionitis rufa</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Jamesonia verticalis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Llavea cordifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Mildella intramarginalis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)



Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Monogramma graminea</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Monogramma paradoxa</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Neurocalis praestantissima</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Onychium japonicum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Onychium siliculosum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Pentagramma triangularis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Pityrogramma austroamericana</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Pityrogramma calomelanos</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Pityrogramma ebenea</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Pityrogramma trifoliata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Platyrama microphyllum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Polytaenium cajenense</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Polytaenium chlorosporum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae	Pteridaceae	Wet ashing technique	<i>Polytaenium guayanense</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae		Wet ashing technique	<i>Polytaenium lanceolatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Polytaenium lineatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Polytaenium urbanii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris arborea</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris argyraea</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris multifida</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris propinqua</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris quadriaurita</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris ryukyuensis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris tremula</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Pteris vittata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Chauhan et al. (2011)
Pteridaceae		Wet ashing technique	<i>Pterozonium brevifrons</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Radiovittaria gardneriana</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Radiovittaria minima</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
Pteridaceae		Wet ashing technique	<i>Radiovittaria remota</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Scoliosorus ensiformis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Vittaria amboinensis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
Pteridaceae		Wet oxidation technique	<i>Vittaria carcina</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
Pteridaceae		Wet oxidation technique	<i>Vittaria elongate</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
Pteridaceae		Wet ashing technique	<i>Vittaria flavicosta</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Vittaria graminifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet oxidation technique	<i>Vittaria himalayensis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
Pteridaceae		Wet ashing technique	<i>Vittaria isoetifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet ashing technique	<i>Vittaria lineata</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Sundue (2009)
Pteridaceae		Wet oxidation technique	<i>Vittaria sikkimensis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
Pteridaceae		Wet oxidation technique	<i>Vittaria zosterifolia</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
Pteridaceae		Wet oxidation technique	<i>Athyrium nigripes</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
Pteridaceae		Wet oxidation technique	<i>Diplazium esculentum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)

(Continued)

Table 1. (Continued).

Plant taxa	Family	Techniques used	Plant	Plant parts	Phytolith type	Reference
	Thelypteridaceae	Wet oxidation technique	<i>Ampelopteris prolifera</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Thelypteridaceae	Wet oxidation technique	<i>Christella dentate</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Thelypteridaceae	Wet oxidation technique	<i>Cyclosorus interruptus</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Thelypteridaceae	Wet oxidation technique	<i>Pronephrium nudatum</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Thelypteridaceae	Wet oxidation technique	<i>Pseudocyclosorus tyloides</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Thelypteridaceae	Wet oxidation technique	<i>Pseudophegopteris pyrrothachis</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Blechnaceae	Wet oxidation technique	<i>Blechnum orientale</i>	Leaf	Elongated epidermal plates with smooth surface and lobed margins	Mazumdar (2011)
	Dryopteridaceae	Wet oxidation technique	<i>Leucostegia immerse</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Dryopteridaceae	Wet oxidation technique	<i>Peranema cyatheoides</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Oleandraceae	Wet oxidation technique	<i>Polystichum squarrosus</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Oleandraceae	Wet oxidation technique	<i>Oleandra wallichii</i>	Leaf	Blunt ends and sides that are smooth, denticulate sparsely denticulate or shallowly lobed	Mazumdar (2011)
	Polypodiaceae	Wet oxidation technique	<i>Arthromeris wallichiana</i>	Leaf	Elongated flat base and a surface with two undulating ridges parallel to each other	Mazumdar (2011)
	Polypodiaceae	Dry ashing technique	<i>Drynaria quercifolia</i>	Leaf	Elongated flat base and a surface with two undulating ridges parallel to each other	Chauhan et al. (2011)
	Polypodiaceae	Wet oxidation technique	<i>Phymatopteris griffithiana</i>	Leaf	Elongated flat base and a surface with two undulating ridges parallel to each other	Mazumdar (2011)
	Zygophyllaceae	Dry ash technique	<i>Creosote</i>	Leaf wood	Globular echinate cubic parralelepipedal	Morgan-Edel, Boston, Spilde, & Reynolds (2015)
	Moraceae	Dry ash technique	<i>Mulberry</i>	Leaf twig	Stomata crenulated tissue cubic	Morgan-Edel et al. (2015)

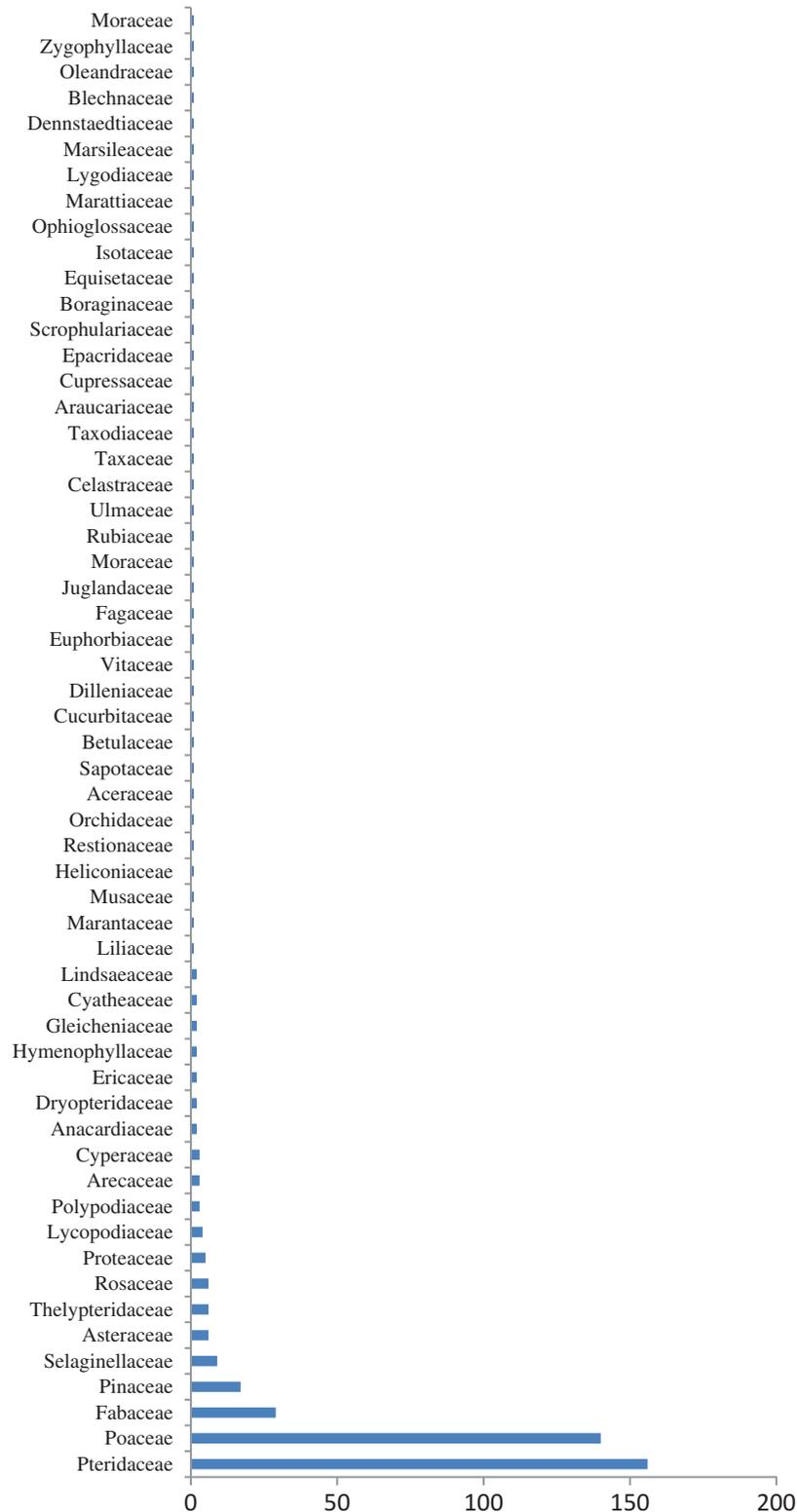


Figure 1. Families in which phytolith analysis have been reported.

climate change. However, modifications with addition of basalt powder indicate good method to enhance the biogeochemical carbon sequestration in crops and may be an effective method to reduce the global warming indirectly (Guo et al., 2015).

Conclusions

The present review attempts to compile the data of phytoliths in various plants analyzed by various researchers from time to time. It will help the workers of scientific community in tracing the relationship

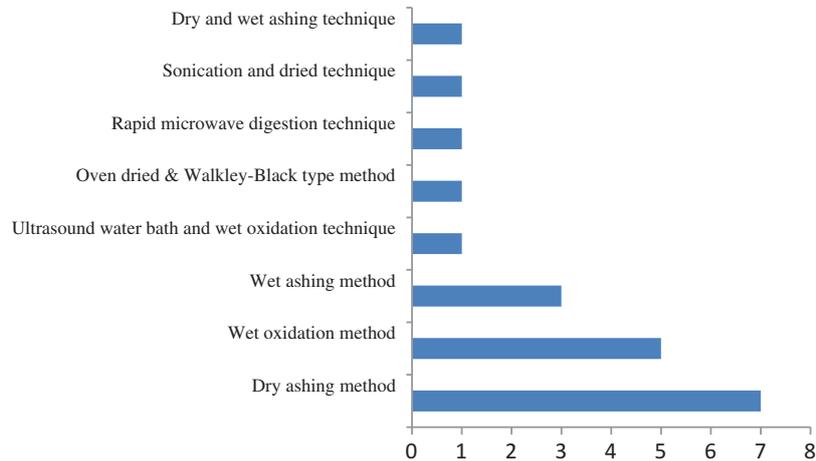


Figure 2. Methodology used by researchers for sample preparation.

between phytoliths and its source plant at the target sites. It was observed that maximum plants eked for phytolith distribution belonged to family Pteridaceae followed by Poaceae family. The most common method used for the digestion of samples for phytoliths preparation is dry ashing method followed by wet oxidation method. The most common type of phytolith reported was blunt ends with smooth followed by rondel and elliptical type of phytoliths.

Disclosure statement

No potential conflict of interest was reported by the authors.

References

- Agarie, S., Agata, W., Uchida, H., Kubota, F., & Kaufman, P. B. (1996). Function of silica bodies in the epidermal system of rice (*Oryza sativa* L.): Testing the window hypothesis. *Journal of Experimental Botany*, 47(5), 655–660.
- Albert, R. M., & Weiner, S. (2001). Study of phytoliths in prehistoric ash layers from Kebara and Tabun caves using a quantitative approach. *Phytoliths: applications in earth sciences and human history*, 251–266.
- Blackman, E., & Parry, D. W. (1968). Opaline silica deposition in rye (*Secale cereale* L.). *Annals of Botany*, 32(1), 199–206.
- Blinnikov, M. S. (2005). Phytoliths in plants and soils of the interior Pacific Northwest, USA. *Review of Palaeobotany and Palynology*, 135(1), 71–98.
- Carnelli, A. L., Madella, M., & Theurillat, J. P. (2001). Biogenic silica production in selected alpine plant species and plant communities. *Annals of Botany*, 87(4), 425–434.
- Carnelli, A. L., Theurillat, J. P., & Madella, M. (2004). Phytolith types and type-frequencies in subalpine–Alpine plant species of the European Alps. *Review of Palaeobotany and Palynology*, 129(1), 39–65.
- Chauhan, D. K., Tripathi, D. K., Kumar, D., & Kumar, Y. (2011). Diversity, distribution and frequency based attributes of phytolith in *Arundo donax* L. *International Journal Innov Biologic Chemical Sciences*, 1, 22–27.
- Chen, C., & Lewin, J. (1969). Silicon as a nutrient element for equisetum arvense. *Canadian Journal of Botany*, 47, 125–131. doi:10.1139/b69-016
- Collura, L. V., & Neumann, K. (2017). Wood and bark phytoliths of West African woody plants. *Quaternary International*, 434, 142–159.
- Epstein, E. (1999). *Annual Review Oof Plant Physiology and Plant Molecular Biology*, 50, 641–644. doi:10.1146/annurev.arplant.50.1.641
- Esteban Alamá, I. (2016). Reconstructing past vegetation and modern human foraging strategies on the south coast of South Africa (Doctoral dissertation). Universitat de Barcelona.
- Geis, J. W. (1973). Biogenic silica in selected species of Gramineae. *Annals of Botany*, 42, 1119–1121. doi:10.1093/oxfordjournals.aob.a085552
- Guo, F., Song, Z., Sullivan, L., Wang, H., Liu, X., Wang, X., & Zhao, Y. (2015). Enhancing phytolith carbon sequestration in rice ecosystems through basalt powder amendment. *Science Bulletin*, 60(6), 591–597.
- Hart, T. C. (2016). Issues and directions in phytolith analysis. *Journal of Archaeological Science*, 68, 24–31.
- Iriarte, J., & Paz, E. A. (2009). Phytolith analysis of selected native plants and modern soils from southeastern uruguay and its implications for paleoenvironmental and archeological reconstruction. *Quaternary International*, 193(1–2), 99–123. doi:10.1016/j.quaint.2007.10.008
- Jones, L. H. P., & Milne, A. A. (1963). Studies of silica in the oat plant: I. Chemical and physical properties of the silica. *Plant and Soil*, 207–220.
- Klein, R. L., & Geis, J. W. (1978). Biogenic silica in the pinaceae. *Soil Science*, 126(3), 145–156.
- Korolúk, E. A. E. Natal'á Viktorovna Polos' mak. (2010). *Plant remains from Noin Ula burial mounds 20 and 31 (Northern Mongolia)*. Elsevier BV.
- Lentfer, C., Therin, M., & Torrence, R. (2002). Starch grains and environmental reconstruction: a modern test case from west new britain, papua new guinea. *Journal of Archaeological Science*, 29(7), 687–698. doi:10.1006/jasc.2001.0783
- Li, B., Song, Z., Li, Z., Wang, H., Gui, R., & Song, R. (2014). Phylogenetic variation of phytolith carbon sequestration in bamboos. *Scientific reports*, 4.
- Lisztes_Szabo, Z. S. U. Z. S. A., Kovács, S., & Pető, Á. (2014). Phytolith analysis of *Poa pratensis* (Poaceae) leaves. *Turkish Journal of Botany*, 38(5), 851–863.

- Ma, J. F., & Yamaji, N. (2006). Silicon uptake and accumulation in higher plants. *Trends in Plant Science*, 11, 392–397. doi:10.1016/j.tplants.2006.06.007
- Matoh, T., Kairusmee, P., & Takahashi, E. (1986). Salt-induced damage to rice plants and alleviation effect of silicate. *Soil Science and Plant Nutrition*, 32, 295–311. doi:10.1080/00380768.1986.10557506
- Mazumdar, J. (2011). Phytoliths of pteridophytes. *South African Journal of Botany*, 77(1), 10–19.
- Mercader, J., Bennett, T., Esselmont, C., Simpson, S., & Walde, D. (2009). Phytoliths in woody plants from the Miombo woodlands of Mozambique. *Annals of Botany*, 104(1), 91–113.
- Morgan-Edel, K. D., Boston, P. J., Spilde, M. N., & Reynolds, R. E. (2015). Phytoliths (plant-derived mineral bodies) as geobiological and climatic indicators in arid environments. *New Mexico Geology*, 37(1), 3–20.
- Novello, A., Barboni, D., Sylvestre, F., Lebatard, A. E., Paillès, C., Bourlès, D. L., & Brunet, M. (2017). Phytoliths indicate significant arboreal cover at Sahelanthropus type locality TM266 in northern Chad and a decrease in later sites. *Journal of Human Evolution*, 106, 66–83.
- Piperno, D. R. (1988). *Phytolith Analysis. An archaeological and geological perspective*, 280. London, UK: Academic Press.
- Piperno, D. R. (2006). *Phytoliths: A comprehensive guide for archaeologists and paleoecologists*. Lanham, MD: AltaMira Press.
- Piperno, D. R., & Pearsall, D. M. (1998). The silica bodies of tropical American grasses: Morphology, taxonomy, and implications for grass systematics and fossil phytolith identification. In *Smithsonian Contributions to Botany No. 85*. Washington, DC: Smithsonian Institution Press.
- Ryan, P. 2014. Phytolith studies in archaeology. In C. Smith (Ed.), *Encyclopedia of global archaeology*. New York: Springer. doi: 10.1007/9781441904652_2258.
- Shakoor, S. A., & Bhat, M. A. (2014). Morphological diversity of phytolith types in some chloridoid grasses of Punjab. *International Journal of Botany and Research*, 4 (1), 1–10.
- Song, Z., Liu, H., Strömberg, C. A., Yang, X., & Zhang, X. (2017). Phytolith carbon sequestration in global terrestrial biomes. *Science of the Total Environment*, 603, 502–509.
- Strömberg, C. A. (2002). The origin and spread of grass-dominated ecosystems in the late tertiary of North America: preliminary results concerning the evolution of hypsodonty. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 177(1-2), 59–75. doi:10.1016/S0031-0182(01)00352-2
- Struve, G. A., 1835. De Silicia in Plantis Nonnullis. Dissertation. Unviersitate Litteraria.
- Sundue, M. (2009). Silica bodies and their systematic implications in Pteridaceae (Pteridophyta). *Botanical Journal of the Linnean Society*, 161(4), 422–435.
- Thorn, V. C. (2008). New Zealand sub-Antarctic phytoliths and their potential for past vegetation reconstruction. *Antarctic Science*, 20(1), 21–32.
- Twiss, P. C. (1987). Grass-opal phytoliths as climatic indicators of the Great Plains Pleistocene. In W. C. Johnson (ed.), *Quaternary Environments of Kansas* (Vol. 5, pp. 179–18). Kansas: Kansas Geological Survey Guide book Series.
- Wu, H., Zhang, S., Ma, Y., Zhou, J., Luo, H., & Yang, J. (2018). Comparison of microbial communities in the fermentation starter used to brew Xiaoqu liquor. *Journal of the Institute of Brewing*, 123(1), 113–120.
- Yang, X., Wu, W., Liu, K., Chen, W., & Zhou, Z. (2018). Multiple dictionary pairs learning and sparse representation-based infrared image super-resolution with improved fuzzy clustering. *Soft Computing*, 22(5), 1385–1398.
- Zurro, D., García-Granero, J. J., Lancelotti, C., & Madella, M. (2016). Directions in current and future phytolith research. *Journal of Archaeological Science*, 68, 112–117.